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Thanks for your continued financial support:

Northeast Area Office (USDA-ARS, NEA) – Dariusz Swietlik (Area Director)

USDA – APHIS/BRS – Bernadette Juarez (Deputy Administrator)

Organization

Organizing Committees: Lots of people provide the support and staffing for this meeting! Many thanks to all of them for the fine job they are doing. If you would like to join a committee and help, please let us know. We are always looking for dedicated volunteers!

Program:

Ben Matthews Jim Saunders John Hammond Ken Haymes Nadim Alkharouf Savi Natarajan Chris Clarke Fred Gouker

Publicity: Ben Matthews Jim Saunders Program booklet: David Puthoff

Web Page: Nadim Alkharouf

Treasurer: Jim Saunders

Poster Organizer: Chris Clarke **Poster Judges:** Srinivasa Chaluvadi Fred Gouker Peter Abrahamian Chris Clarke

Audio-Visual / Technology Assistance: Nadim Alkharouf

Registration: Nadim Alkharouf Jim Saunders

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2020 MAPMBS 37th Annual Meeting Schedule - Virtual

Wednesday, August 19, 2020

9:00 Welcome - Meeting Information - Technology/Etiquette - John Hammond

9:05 - 9:40 -N. Bhushan Mandava **Repar Corporation Brassinosteroids:** Sixth Group of Plant Hormones

Introduction: Viji Sitther

9:40 - 10:15 Wei Wei **USDA-ARS, MPPL** Data from -omics studies indicate phytoplasma is capable of manipulating host metabolism in favor of its own survival.

Introduction: John Hammond

10:15 - 10:30 Posters; Live Summary, and Chat (details to come)

10:30 - 10:45 Break

10:45 – 11:20 James Kim **PPGR-USDA-ARS** Effective Use of High Throughput Phenotyping for Plant Breeding Introduction: John Hammond

11:20 – 11:55 Justin Overcash **APHIS USDA** Engineered Insects and Co.: Applications for Plant Protection

Introduction: Ken Haymes

- 11:55 12:15 Posters; Live Summary, and Chat (details to come)
- 12:15 1:30Lunch

1:30 - 2:05**Rubella Goswami Branch Chief USDA-APHIS** Insights into funding programs and tips for grant writing Introduction: Ken Haymes

2:05 - 2:40University of Massachusetts **Madelaine Bartlett** Using phylogenetic analysis to find the dark matter of genomes: microproteins and more Introduction: John Hammond

- 2:40 3:00 Break - Posters; Live Summary, and Chat (details to come)
- 3:00 3:15**Announcement of Poster Contest Winners**
- 3:15 3:20 **Introduction of Keynote speaker: Fred Gouker**
- 3:20 4:20The Leslie Wanner Keynote speaker: **Markita Landry** University of California, Berkeley Nanomaterials Enable Delivery of Genetic Material Without Transgene Integration in Mature Plants

4:20 Thanks for "attending" - John Hammond

BRASSINOSTEROIDS: SIXTH GROUP OF PLAT HORMONES

N. Bhushan Mandava

Repar Corporation, 8070 Georgia Avenue, Silver Spring, Maryland 20910, USA

Plant hormones play a crucial role in plant growth and development. There were originally 5 major plant hormones, auxins, gibberellins, cytokinins, ethylene and abscisic acid. All these hormones have independent roles within the plant, but also work in tandem in order to regulate plant growth and development. As such, commercial versions of these plant growth hormones [also referred to as plant growth regulators (PGRs)] were developed for their application to agricultural crops.

The first discovery of new plant hormones, named Brassins, was reported by Mitchell and Mandava in 1970 at USDA Laboratory in Beltsville, Maryland. Over a 9-year period at USDA Laboratories in Beltsville, Philadelphia, Peoria, Dr. N. Bhushan Mandava and his coworkers were involved in identifying the active substance from Brassins. He, along with a team of other chemists, biochemists and chemical engineers developed a chemical process that isolated 10 mg of a pure crystalline product from over 500 lbs of rape (Brassica napus) pollen. The discovery of this substance, later named Brassinolide (present at 200 ppb level), is considered one of the greatest scientific achievements in plant sciences. It is recognized as the SIXTH major plant hormone group. and has major cellular implications in the realm of plant biology. In 1979, Dr. Grove and Dr. Mandava characterized it as brassinolide, which is the first plant hormone in a group of over 70 brassinosteroids found in nature. Since brassinolide is difficult to synthesize, the closely related analogs epi-brassinolide (EBR) and 28-homobrassinolide (HBR) were synthesized and developed for commercial uses. Besides Brassinolide (BL), HBR and EBR are the most active brassinosteroids. In the U.S., Switzerland and India, HBR is registered for commercial uses. Like BL, HBR enhances cell division and elongation, interacts synergistically with other other Plant Hormones, and protects plants from a variety of stress factors such as water, salt, heat, etc. It also elicits profound physiological responses at sub-micromolar concentrations. This is highly significant because farmers will need to use less material overall to reap the application benefits, which leads to more sustainable usage. The yield increases are verified by multiple field studies conducted in the United States and South America (Chile) as well as in Switzerland and India.

Application of HBR shows the disease and climate resistance properties, which can further help growers achieve higher yields. HBR also perpetuates substantial yield increase in nut crops, with similar results in trialing with almonds, and walnuts. Dr. Mandava group applied for the registration of Homobrassinolide (HBR) with the U.S. Environmental Protection Agency (EPA) in order to commercialize HBR. The EPA granted the registration for HBR technical as a biopesticide in 2010 after reviewing the pertinent documentation regarding product chemistry, toxicology and other safety data. The EPA also granted a tolerance exemption for HBR, which means that the residues in HBR in treated food and fiber commodities are not of any health and safety concerns, and that HBR can therefore be used on all crops. To further prepare the product for a global market, the usage of HBR on fruit and nut crops (Grapes, Almonds and Walnuts) was also patented, with additional patents being developed for other crops. As a next generation PGR, HBR and other brassinosteroids represent the latest agrochemical technology, and will be increasingly implemented across the world. Currently, there are ongoing initiatives to bring HBR into the EU (starting with Switzerland), Thailand, Kenya, and more.

DATA FROM OMICS STUDIES INDICATE PHYTOPLASMA IS CAPABLE OF MANIPULATING HOST METABOLISM IN FAVOR OF ITS OWN SURVIVAL

Wei Wei1, Yan Zhao1, Yue Tan2, and Qingzhong Liu2

1Molecular Plant Pathology Laboratory, Beltsville Agricultural Research Center, Agricultural Research Service, United States Department of Agriculture, Beltsville, MD, USA 2Shandong Institute of Pomology, Taian, China

Phytoplasmas are small, wall-less plant pathogenic bacteria that cause various diseases worldwide and adversely impact agricultural economy. During the course of infection, the metabolic networks between phytoplasmas and their plant hosts are interlinked. Phytoplasmas and hosts compete for similar or identical nutrient substrates within the microenvironment, and as a result, a slight alteration in metabolism could significantly affect the outcome of phytoplasma-host interactions. Genomics studies have disclosed that phytoplasmas are highly host dependent, because phytoplasmas have undergone reductive evolution, and lost many genes that are involved in metabolic pathways essential for free-living organisms. Therefore, phytoplasmas must modulate the metabolism of host plant cells for the supply of nutrients, energy, and metabolites to establish successful replication and infection in plants. By employing metabolomics approach and integrating existing omics data, the present study revealed that phytoplasma infection promoted glycolysis and increased pentose phosphate pathway (PPP) activities in plants. Enhanced glycolysis and PPP activities not only provided energy and nutrient substances, but also facilitated biosynthesis of necessary low molecular metabolites, including amino acids, nucleotides and fatty acids/lipids, which are conducive to phytoplasma replication and infection. Conceivably, the host plant relies on the similar nutrient substrates and low molecular metabolites within the same microenvironment to support host defense response to phytoplasma infection. A prime example is enhanced flavonoid biosynthesis upon phytoplasma infection. Phytoplasma infection also led to the accumulation of a compound that attracts phloem sap-sucking insects, which is beneficial to the survival and transmission of phytoplasma. These findings indicate that phytoplasma can induce metabolic reprogramming in host plants to favor its own growth and infection.

EFFECTIVE USE OF HIGH THROUGHPUT PHENOTYPING FOR PLANT BREEDING

James Y. Kim

Plant Physiology and Genetics Research Unit;USDA-ARS, US Arid-Land Agricultural Research Center

Plant breeding and biotechnology are a game changer of ag industry to meet food demand in next decades. Phenotyping is critical to capture genotypic traits of plants through characterizing the plant responses to abiotic stresses such as heat and drought in Arizona. High throughput phenotyping (HTP) plays a key role in field mapping and data processing for timely advancement decisions. The seminar presents why HTP is important and how the HTP system is designed and implemented, as well as analytics software to visualize and analyze the phenotypic data. Discussion includes scale-up and technology transfer for small\large-scale plant sensing.

ENGINEERED INSECTS & CO.: APPLICATIONS FOR PLANT PROTECTION

Justin Overcash USDA-APHIS

There is great interest in developing novel techniques and strategies for pest management. Biotech innovations such as engineered insects and microbes could complement an existing toolbox for plant protection by reducing pesticide use and countering resistant pests and pathogens. The discovery and application of CRISPR/Cas9 and associated technologies has stimulated research into various ways to control pest populations. Furthermore, RNAi-based techniques may also provide solutions to emerging and existing impacts by pests and pathogens. In this talk I will present a sampling of recent developments in genetically manipulating insects and microbes for plant protection.

INSIGHTS INTO FUNDING OPPORTUNITIES AND TIPS FOR GRANT WRITING

Rubella S. Goswami,

Branch Chief USDA-Animal and Plant Health Inspection Service; Former National Program Leader, USDA-National Institute of Food and Agriculture

This presentation will provide an overview about the federal funding programs administered by USDA-National Institute of Food and Agriculture (NIFA), the primary extramural funding arm of USDA that supports both capacity and competitive programs. It will highlight some programs relevant to those working in the area of plant sciences that support applied and basic research as well as pre and post-doctoral fellowships. Tips for accessing information regarding funding opportunities and successful grant writing will also be shared.

COMPLEX FAMILIES, COMPLEX HISTORIES: EVOLUTION AND DIVERSIFICATION OF THE LEUCINE RICH REPEAT -RECEPTOR-LIKE KINASE (LRR-RLK) GENE FAMILY

Jarrett Man, Joseph P. Gallagher, Madelaine Bartlett Biology Department, UMass Amherst, 611 North Pleasant Street, Amherst, MA mbartlett@umass.edu

LRR-RLK proteins mediate cell-cell signaling in plant development and defense. For example, LRR-RLKs in the CLAVATA1 (CLV1) clade are essential regulators of meristem homeostasis and, in turn, the development of plant form. The LRR-RLK gene family has diversified to an extreme degree; both through whole genome and smaller-scale duplications. The immense size and complexity of the LRR-RLK gene family has made resolving gene trees challenging. To address this challenge, we developed an iterative search and tree-building method for capturing all the LRR-RLKs and related proteins encoded in a genome, and for resolving the evolutionary history of the gene family. We discovered numerous instances of domain loss; domain gain through duplication and subsequent gene fusion events; and domain swapping between LRR-RLK clades. Moving forward, we are using the tools available in Arabidopsis thaliana to assess the functional evolution of developmental regulators in the LRR-RLK gene family. Our fine-grained history of these important genes sets the stage for understanding the diversification of signaling in the evolution of plant development and defense.

NANOMATERIALS ENABLE DELIVERY OF GENETIC MATERIAL WITHOUT TRANSGENE INTEGRATION IN MATURE PLANTS

Gozde S. Demirer,^{a,c} & Markita P. Landry,^{a,b,c,d,e a}

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Genetic engineering of plants is at the core of sustainability efforts, natural product synthesis, and agricultural crop engineering. The plant cell wall is a barrier that limits the ease and throughput with which exogenous biomolecules can be delivered to plants. Current delivery methods either suffer from host range limitations, low transformation efficiencies, tissue regenerability, tissue damage, or unavoidable DNA integration into the host genome. Here, we demonstrate efficient diffusion-based biomolecule delivery into tissues and organs of intact plants of several species with a suite of pristine and chemically-functionalized high aspect ratio nanomaterials. Efficient DNA delivery and strong protein expression without transgene integration is accomplished in mature Nicotiana benthamiana, Eruca sativa (arugula), Triticum aestivum (wheat) and Gossypium hirsutum (cotton) leaves and arugula protoplasts. Notably, we demonstrate that transgene expression is transient and devoid of transgene integration into the plant host genome. We demonstrate that our platform can be applied for CRISPR-based genome editing for transient expression of Cas9 and gRNAs. We also demonstrate a second nanoparticlebased strategy in which small interfering RNA (siRNA) is delivered to mature Nicotiana benthamiana leaves and effectively silence a gene with 95% efficiency. We find that nanomaterials both facilitate biomolecule transport into plant cells, while also protecting polynucleotides such as RNA from nuclease degradation. DNA origami and nanostructures further enable siRNA delivery to plants at programmable nanostructure loci, which we use to elucidate force-independent transport phenomena of nanoparticles across the plant cell wall. Our work provides a tool for species-independent, targeted, and passive delivery of genetic material, without transgene integration, into plant cells for diverse plant biotechnology applications.

EVOLUTION OF HOST SUSCEPTIBLITY IN THE GOLOVINOMYCES LATISPORUS-ASTERACEAE SYSTEM

Michael Bradshaw and Patrick Tobin

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The host range and severity of pathogens are dependent on interactions with their hosts and are hypothesized to have evolved as products of a coevolutionary arms race. An understanding of the factors that affect host range and pathogen severity is especially crucial in introduced pathogens that infect evolutionarily-naïve hosts and cause substantial damage to ecosystems. Powdery mildews are detrimental pathogens found worldwide in managed and natural systems. Golovinomyces latisporus is a powdery mildew species that is especially damaging to plants within Asteraceae, and in particular plants within Helianthus. In this study, we evaluated 126 species within Asteraceae to measure the role of host plant morpho-physiological traits and evolutionary history on their susceptibility and severity to G. latisporus. We observed a phylogenetic signal in both susceptibility and severity between and within major clades of the Asteraceae. In general, there was a major phylogenetic structure of host severity to G. latisporus, however, there was some fine-scale phylogenetic variability. Phylogenetic statistical methods showed that chlorophyll density, biomass, stomatal index and trichome density were not associated with disease severity, thus providing evidence that phylogenetic structure, rather than observed plant morpho-physiological traits, is the most reliable predictor of pathogen severity. This work sheds light on the role that evolutionary history plays in plant susceptibility and severity to disease and underscores the relative unimportance of commonly assessed host plant traits in powdery mildew severity.

CLONING ospA FOR PLANT (ARABIDOPSIS) EXPRESSION

Eleanor Browne, David Puthoff

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Borrelia burgdorferi (Lyme disease) is a vector-borne disease that is both very common and capable of causing long term negative health effects. Because it is spread by deer ticks, which also bite and transfer infection through mice, our goal is to design an oral vaccine capable of preventing mice from becoming infected. This strategy will use ospA from *B. burgdorferi*, produced by altered Arabidopsis thaliana which will, when eaten, deliver the ospA to mice who will begin to produce antibodies effective against B. burgdorferi. Primer pairs for both control and ospA amplification were designed Both primer pairs amplified the respective fragments which have been kept refrigerated due to a laboratory shutdown due to COVID-19. The ospA coding sequence will be expressed in Arabidopsis using pCambia 1201 The cloned ospA/ pCAMBIA plasmid will be transferred into Agrobacterium tumefaciens to be transferred to A. thaliana by floral dip. The offspring of these dipped A. thaliana will be grown, tested for expression of ospA and crossed for homozygosity of the transgene. Those which express ospA at the highest level will be used in tests with mice which will be further tested to determine whether the mice have developed the necessary antibodies to have protection against *B. burgdorferi*.

FUNCTIONAL CHANGES OF DNA METHYLATION PATTERNS OVER MICRORNA GENES DURING SOYBEAN NODULE FORMATION AND DEVELOPMENT

Valeria S. Lopes-Caitar1, ‡, Sarbottam Piya1,‡, Won-Seok Kim2, Vince Pantalone1, Hari B. Krishnan2,3, Tarek Hewezi1,*

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DNA methylation is a widely spread epigenetic mark that contributes to gene regulation in various developmental programs, including nodulation. Nevertheless, it is currently unknown whether microRNA (miRNA) genes are subjected to DNA methylation changes that may impact their expression, and hence their function during nodule development in soybean (Glycine max). In the current study, we examined DNA methylation levels and patterns of miRNA genes at three distinct nodule developmental stages (formation, development and senescence) using wholegenome bisulfite sequencing (BS-seq) and RNA-seq data. Bulk DNA methylation levels over miRNA genes (promoter and primary transcript regions) in the CG, CHH and CHH sequence contexts were significantly higher in the nodules than the corresponding control root tissues. A significant number of differentially methylated miRNAs were identified mainly in the promoter region and to a much lesser extent in the primary transcript region. Quantification of miRNA expression levels revealed a link between DNA methylation in the promoter and primary transcript regions, and miRNA expression and biogenesis, respectively. RNA-seq data analysis revealed that the expression patterns of miRNA target genes were associated with the methylation status of their negative regulators, particularly during development and senescence stages. More specifically, target genes of differentially hypomethylated miRNAs were downregulated, whereas target genes of differentially hypermethylated miRNAs were up-regulated. Taken together, our data demonstrate that DNA methylation of miRNA genes is of functional significance and may contribute nodule transcriptome reprograming during development and senescence.

DNA METHYLATION PATTERNS DURING EMBRYOGENESIS AND SEED DEVELOPMENT IN ARABIDOPSIS

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DNA methylation and demethylation precisely and effectively modulate gene expression during plant growth and development and in response to fluctuating environments. Recent experiment evidence indicates that patterns and levels of DNA methylation are dynamically reprogramed during embryogenesis and seed development. However, expression profiles of genes involved in DNA methylation and demethylation during embryogenesis remain partially known. Thus, we have characterized the spatiotemporal expression patterns of genes involved in DNA methylation and active demethylation in developing seeds of Arabidopsis using GUS reporter lines. Our data shows the expression of several genes throughout seed development, indicating that the methylomes of developing seeds are established through coordinated interactions between DNA methylation and demethylation pathways. Transcriptional activity of the CG methyltransferase MET1 and its homologs MET2b, MET3 was detected in all embryogenesis stages assessed. Our expression analysis also revealed that the non-CG methyltransferases CMT1 and CMT2 were expressed in a sequential manner during embryogenesis and seem to support the RdDM pathway methyltransferase DRM2 in establishing de novo CHH methylation sites. In addition to confirming the key role of DME in mediating extensive hypomethylation in the endosperm, our suggests DML2 and DML3 aid in this process. Taken together, our results demonstrate that DNA methylation pathways act in an overlapping and specific manner in various seed and embryonic tissues, highlighting the dynamic configuration in the patterns and levels of DNA methylation during seed development.

EVALUATION OF A NEW METHOD FOR DNA EXTRACTION OF "LEGAL HIGH" PLANT SPECIES

Cassandra P. O'Hern, B.S., Angelique L. Ryan, B.S., and Kelly M. Elkins, Ph.D. Towson University, Chemistry Department, Forensic Science Program, 8000 York Rd, Towson, MD 21252 kmelkins@towson.edu

As of late, the investigation of plant species that produce "legal highs", or chemicals, plants, or fungi accessible without legal restrictions that mimic the psychoactive effects of structurally similar controlled substances, have become increasingly relevant within the field of forensic science. Consequently, it is important to develop a straightforward, high yield, and practical nucleic acid isolation process to integrate into a forensic laboratory's operating procedures. The focus of this study was placed on the capability of the MicroGEM PDQeX phytoGEM system, which utilizes enzymatic reactions and an innovative cartridge structure that degrades the cell wall and purifies DNA, to extract amplifiable DNA from several known legal high plant species: Artemisia absinthium, Datura stramonium, Ipomoea purpurea, Mitragyna speciosa, and Papaver somniferum. The pulverized plant material DNA was extracted using the PDQeX Nucleic Acid Extractor plant protocol, quantified using the Invitrogen Qubit 2.0 Fluorometer, and evaluated for amplifiability by PCR-HRM using the Qiagen Rotor-Gene Q. The melt curves produced from this method of extraction were concordant with those examined in a previous study that utilized the Qiagen DNeasy Plant Mini Kit for extraction. The PDQeX phytoGEM system proved to be simple, rapid, affordable, and compatible with downstream processes.

FLOWERING LOCUS T CHIMERIC PROTEIN INDUCES FLORAL PRECOCITY IN EDIBLE CITRUS

Judith P. Sinn1, Jeremy Held1,2,#, Chad Vosburg1, Sara M. Klee1,3, Vladimir Orbovic4, Earl Taylor5, Tim Gottwald5, Ed Stover5, Gloria Moore6, and Timothy W. McNellis1, * 1Department of Plant Pathology and Environmental Microbiology, The Pennsylvania State University, University Park, PA, USA

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Breeding of perennial tree crops is challenging due to a multi-year juvenility period. Rapid cycle breeding represents a solution by inducing early flowering through constitutive expression of flower-promotion genes. This approach has so far failed in edible citrus varieties due to difficulty regenerating plants overexpressing floral regulatory genes, possibly due to excessive responsiveness to the floral signal. In the present study, we expressed a translational fusion of the Poncirus trifoliata (trifoliate orange) Flowering Locus T (PtFT1) floral regulator to a single-chain variable fragment antibody (scFv) in the edible grapefruit (Citrus x paradisi) cultivar 'Duncan'. We hypothesized that fusion of PtFT to the scFv might attenuate PtFT activity and allow regeneration of precocious plants. Fifteen successful transformants were obtained and grafted onto Carrizo citrange rootstocks. Western blotting revealed variability in the expression levels of PtFT1-scFv protein among the 15 lines. Of the seven lines closely monitored for a flowering phenotype, four displayed precocious flowering within the first year after transfer to a growth chamber. Flowers and inflorescence architecture were morphologically normal in all precocious lines. Fruit developed normally on the PtFT1-scFv plants, but hand pollination was necessary for seed set. A reduction in thorn size was also associated with PtFT1-scFv expression, a trait corresponding to maturity. The highest expressing PtFT1-scFv lines exhibited dwarfism and a weeping growth form. These results document successful constitutive FT expression in edible citrus for the purpose of reducing juvenility and provide a tool for advancing rapid cycle breeding in citrus.

EXPANDING THE SCOPE OF PLANT GENOME ENGINEERING WITH NOVEL CAS12A ORTHOLOGS AND HIGHLY MULTIPLEXABLE EDITING SYSTEMS

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CRISPR-Cas12a is a promising genome editing system for targeting AT-rich genomic regions. Compared to Cas9, Cas12a has shown higher targeting specificity in plants. Unlike Cas9 that usually generates small deletions, Cas12a cleavage generates staggered ends resulting in larger deletions, making it a suitable nuclease for gene knockout. Moreover, Cas12a only requires a short CRISPR RNA (crRNA) for each target and possesses RNase activity for crRNA array processing, making it an ideal platform for multiplexed editing. Cas12a has been widely applied in plants and achieved high editing efficiencies for single gene targets. However, comprehensive genome engineering using CRISPR usually requires simultaneous targeting of multiple genes at defined locations, which cannot be achieved easily and efficiently with current Cas12a systems, due to their strict PAM (protospacer adjacent motif) requirements and limited multiplex capacity. To expand the targeting scope of Cas12a, we screened nine new Cas12a orthologs and identified six that possess high editing activity and specificity in rice. Among them, Mb2Cas12a stands out with high editing efficiency, relaxed PAM requirements and tolerance to low temperatures. Engineered Mb2Cas12a can also target altered PAMs. These new Cas12a systems have greatly expanded the targeting scope of Cas12a in major crops. To further enable large-scale genome engineering, we compared 12 multiplexed Cas12a systems and identified a potent system that exhibited nearly 100% biallelic editing efficiency with the ability to target as many as 14 sites in rice. This is the highest level of multiplex edits in plants to date using Cas12a. This study has greatly expanded the targeting scope of Cas12a for crop genome engineering.

OVER-EXPRESSION OF CELLULASE ENZYMES IN STREPTOMYCES TO PROBE THE TRANSITION FROM SAPROPHYTE TO PHYTOPATHOGEN

Kara LeClair, Dr. Christopher Clarke, and Chad Kramer USDA, GIFVL, 10300 Baltimore Ave, Beltsville, MD 20705 karaleclair97@gmail.com

Common scab is a disease caused by multiple species of Streptomyces bacteria, resulting in raised or pitted lesions on the surface of potato tubers. Production of the phytotoxin thaxtomin, the main virulence factor produced by pathogenic Streptomyces, is elicited by the disaccharide cellobiose. Therefore, the environmental presence of cellobiose is considered the primary trigger for the transition of pathogenic Streptomyces from saprophyte to pathogen. We are testing the hypothesis that Streptomyces controls its own access to cellobiose through expression of cellulase enzymes that can degrade environmentally ubiquitous cellulose into cellobiose. We identified 35 Streptomyces cellulases and cloned 11 genes into an overexpression vector. We will grow Streptomyces overexpressing cellulase enzymes in the presence of cellulose and observe for elicitation of thaxtomin production. We hypothesize a subset of enzymes in a secreted complex will elicit thaxtomin production. We aim to identify the necessary genes for cellulase production to identify the initial steps of Streptomyces transition from saprophyte to pathogen.

IDENTIFICATION OF PROTEIN KINASE HUB GENES THAT REGULATE SOYBEAN RESISTANCE TO SOYBEAN-CYST NEMATODE

Sarbottam Piya, Bhoomi Patel, Tracy Hawk, and Tarek Hewezi* Department of Plant Sciences, University of Tennessee, Knoxville, TN, 37996 * thewezi@utk.edu

Soybean cyst nematode (SCN, Heterodera glycines) is one of the most devastating pests affecting soybean production worldwide. SCN induces specialized multinucleate feeding site, termed syncytium, in the root vascular tissues. The development of a functional syncytium involves massive induction of signal transduction pathways that orchestrate various cellular functions and activities required for correct cell fate differentiation of this unique cell type. The goal of this study was to investigate the role of protein kinases, which constitute the core components of signal transduction pathways, in soybean-SCN interaction. We generated two independent co-expression networks of soybean genes using control and stress response gene expression data from publicly available RNA-seq datasets. We identified 145 protein kinase hub genes that were highly interconnected in control condition network compared to the stress condition network. Similarly, we identified 247 kinase hub genes that were highly interconnected in stress condition network compared to the control condition network. Out of these 392 protein kinase hub genes, 90 genes were previously identified as differentially expressed in the SCNinduced syncytium, suggesting that these protein kinases may play role in regulating signal transduction pathway during SCN infection. To test the biological significance of the syncytiumkinase hub genes, we assayed SCN susceptibility of soybean plants overexpressing a set of the identified protein kinase hubs using transgenic soybean hairy root system. Overexpression of inactive variants of a set of the kinase hub genes significantly altered soybean susceptibility to SCN. These results suggest that protein kinases are promising targets for developing robust resistance against SCN.

HEMP (CANNABIS SATIVA L.) STRAINS GENETIC COMPOSITION AND CHEMICAL PRODUCTION

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In the United States, the Hemp Farming Act of 2018 nationally legalized hemp (Cannabis sativa L.) for research and cultivation. The Cannabis sativa species yields strains such as marijuana, industrial hemp, and medical hemp; medical hemp is the focus of this study. Hemp's chemical composition varies throughout the plant's tissue. The desired chemicals for medicinal hemp, CBD (cannabidiol) and terpene compounds, are found in high concentrations in the female flowers. In 2019, the USDA set out regulations for medicinal hemp to have no more than 0.3% total THC (tetrahydrocannabinol) concentration. This study is being completed to learn more about hemp's requirements for high CBD production in Maryland. Our research question is: What combination of variety and location results in the best production of chemical compounds in hemp? The study will address variety, location, planting date, and genetic diversity to determine what combinations of variables lead to the best productions of chemical compounds in hemp. Hemp growers located across western Maryland provided the field sites and hemp was sampled when harvestable. Hemp flowers were collected to determine chemical compositionincluding THC, CBD, and terpene content, while the leaves were collected for DNA analysis. Several standard Cannabis sativa microsatellites, along with novel microsatellites, will be used. Analysis is still being completed. With this research, medical hemp growers in Maryland will be able to determine the strains with the best combination of CBD, THC, and terpenes.

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